

Figure 1. Top: triplex resulting from homopurine ligation fragments bound to pyrimidine bases of the circular DNA template (**R** represents backbone of ligating fragments). Note the use of 5-methylcytidine on the Hoogsteen side of the circular template. Bottom: ribbon graphic of a ligation reaction of two ODNs directed by a circular DNA template.

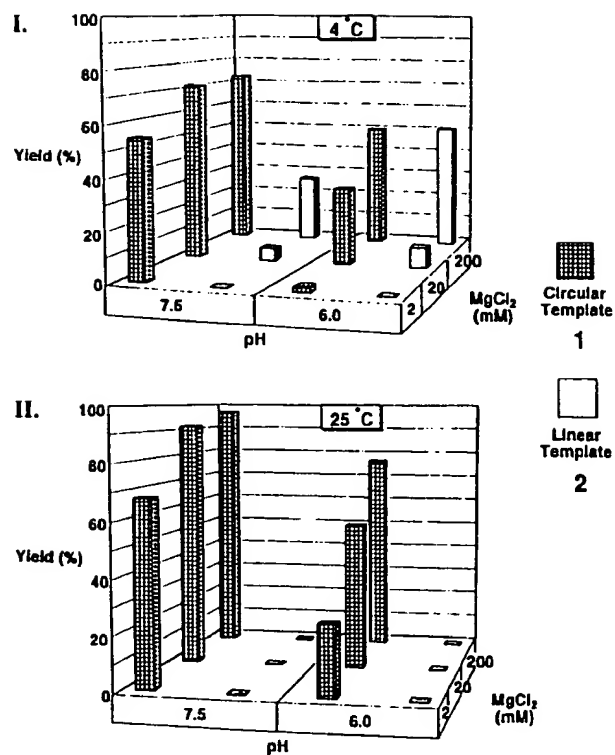


Figure 2. 3-D bar graphs showing the yield (%) of ligation product C. Graph I shows ligation results at 4°C, pH 7.5 and 6.0 with MgCl₂ concentrations of 2, 20 and 200 mM. Graph II shows the same ligation reactions run at 25°C. Data for these graphs was obtained at a reaction time of 3 h. All reactions were reproduced at least twice to afford a % yield error of ± 3 .

Figure 3

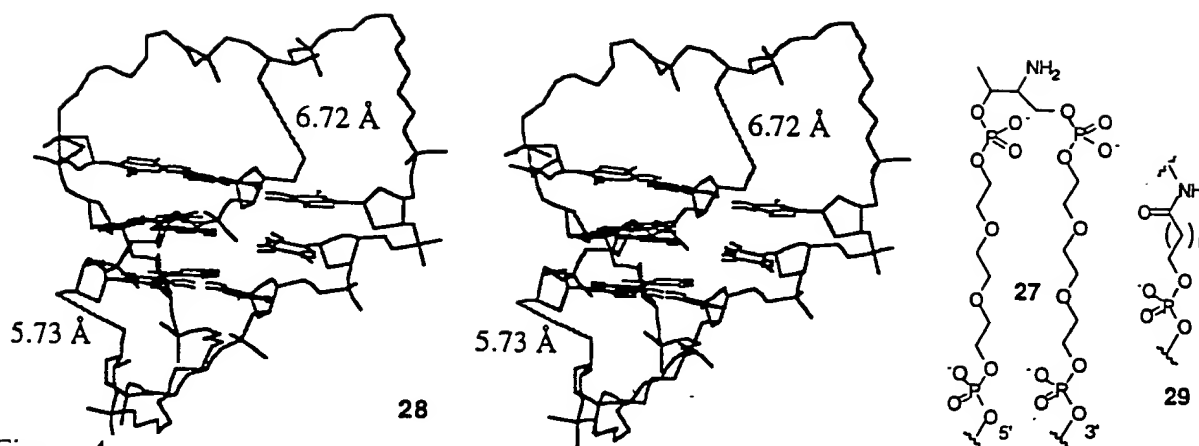
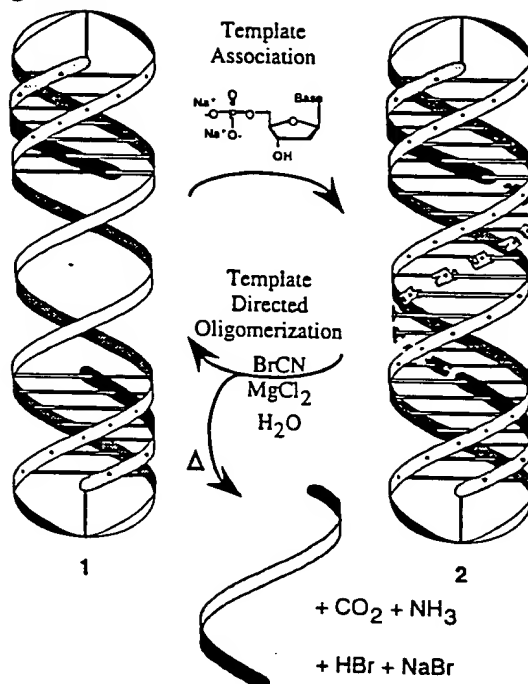
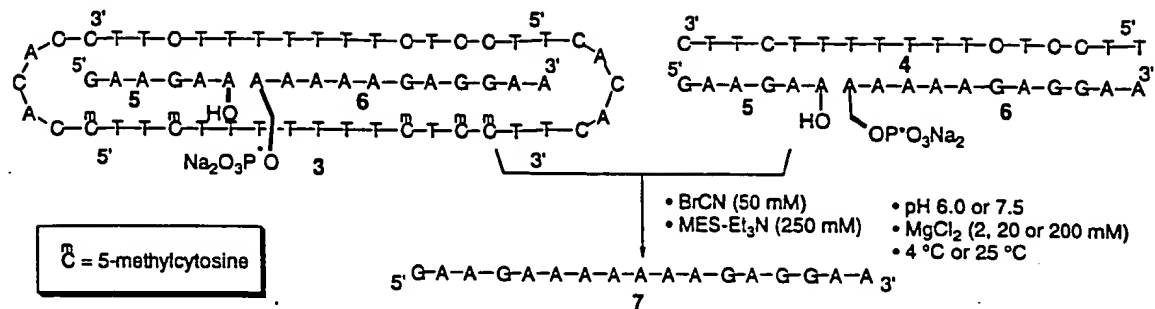


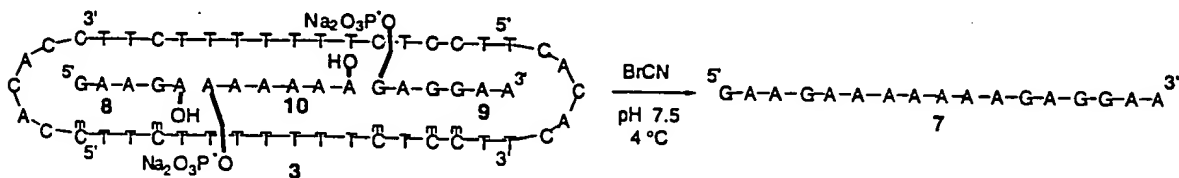
Figure 4 . Stereoview of the energy minimized⁵⁶ circular DNA template (28) with looped linker 27 attached at both ends of the truncated triplex. Based on structure 28, the initial template-primer linker for investigation will be 29 ($n = 2$). Based on an MM2 minimized structure, 29 will span a distance up to 7 \AA .

Scheme 2.

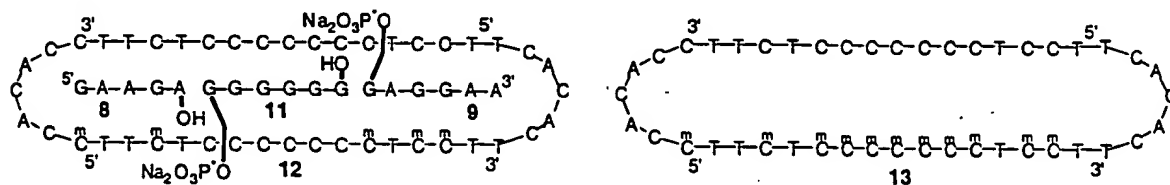


$\overline{\text{C}}$ = 5-methylcytosine

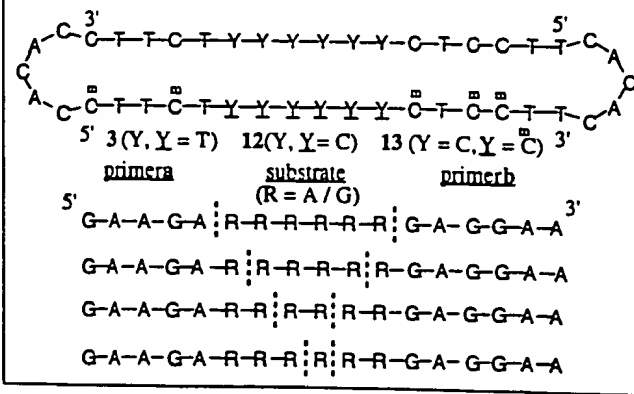
Scheme 3.



Scheme 4.



Scheme 5.



Scheme 1 illustrates the synthesis of the 14-mer DNA template. The process begins with a linear DNA precursor (14) containing a triethoxycarbonyl (Treoc) group at the 3' end. This precursor undergoes directed cyclization to form a circular DNA template (14). The template is then used for the synthesis of a 16-mer DNA strand (16) via a two-step process: (i) reaction with a PEG-protected nucleotide (15) and (ii) reaction with a PEG-protected nucleotide (15) in the presence of DCC and DMF. The final product (16) is a circular DNA strand with a PEG-protected nucleotide at the 3' end.

5' 3 (Y, Y = T) 12 (Y, Y = C) 13 (Y = C, Y = C) 3'

5'	5	substrate	9	3'	ligations
G-A-A-G-A	R-R-R-R-R-R-R	G-A-G-G-A-A	2		
G-A-A-G-A	R-R-R-R-R-R-R	G-A-G-G-A-A	3		
G-A-A-G-A	R-R-R-R-R-R-R	G-A-G-G-A-A	4		
G-A-A-G-A	R-R-R-R-R-R-R	G-A-G-G-A-A	oligomerization		

[illegible]

5'	23	substrate										24	3'	ligations
G-G-G-A-G-G-A-A-A-G-A-A-G	A-A-A-A-A-A-A-A-A-A-A-A	G-G-G-A-A-G-A-G-A-G-G-A	2											
G-G-G-A-G-G-A-A-A-G-A-A-G	A-A-A-A-A-A-A-A-A-A-A-A	G-G-G-A-A-G-A-G-A-G-G-A	3											
G-G-G-A-G-G-A-A-A-G-A-A-G	A-A-A-A-A-A-A-A-A-A-A-A	G-G-G-A-A-G-A-G-A-G-G-A	5											
G-G-G-A-G-G-A-A-A-G-A-A-G	A-A-A-A-A-A-A-A-A-A-A-A	G-G-G-A-A-G-A-G-A-G-G-A	7											
G-G-G-A-G-G-A-A-A-G-A-A-G	A-A-A-A-A-A-A-A-A-A-A-A	G-G-G-A-A-G-A-G-A-G-G-A	oligomerization											

[illegible]

1) Standard amidite additions along with 31, 32 or 33
 2) Deprotection and cleavage
 3) Purification
 3) Triplex directed circularization

3' 30 34

G-C-G-T-C-C-T-T-T-C-T-T-T-T-(Y)₁₂-C-G-G-T-T-G-T-C-T-C-C-T 5'
 X-NH-Treoc H₂N-X
 5' 3'

34 12

31 32 33

DMTO NCEIO NCEIO DMTO
 OEtCN N(iPr)₂ N(iPr)₂ N(iPr)₂ TMS
 ODMT NH-CH₃ NH-CO-O-TMS

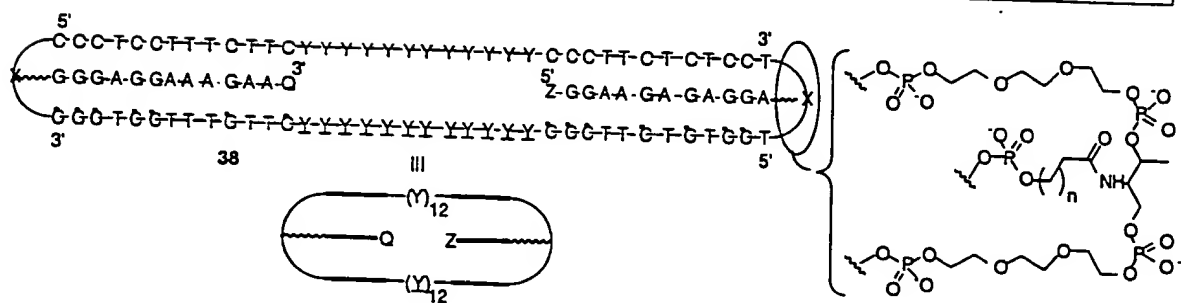
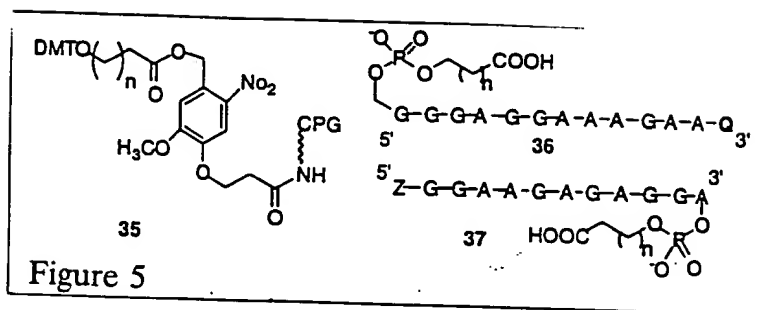
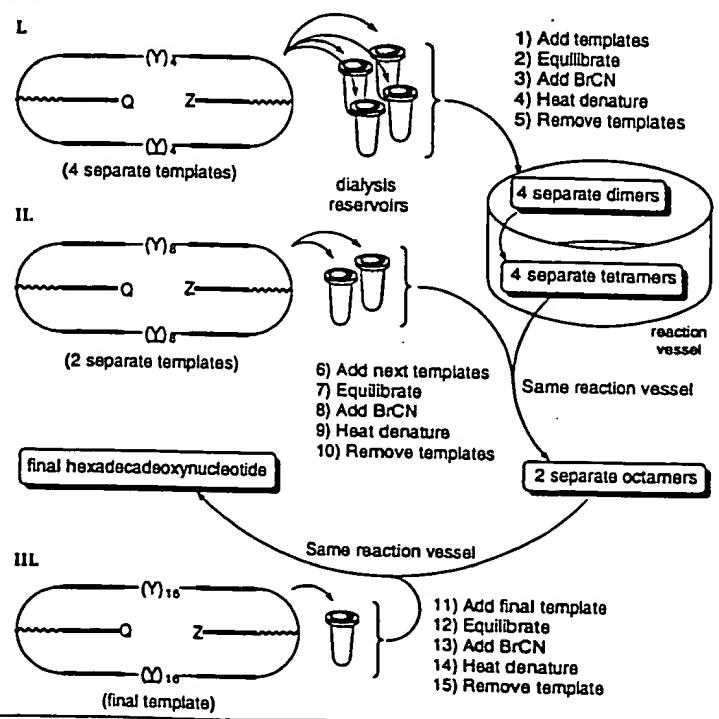
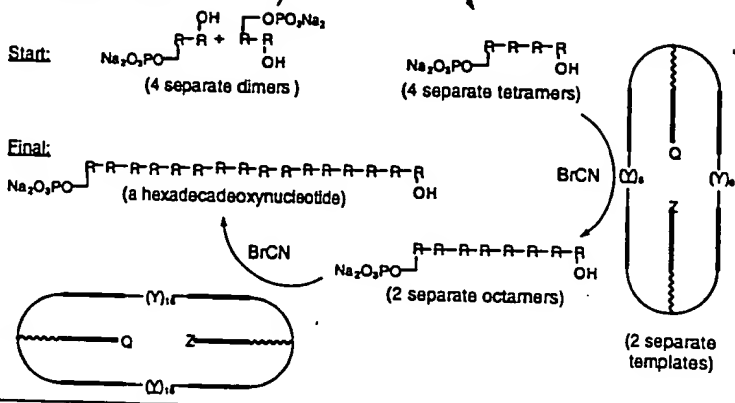


Figure 6 . Primer attached circular DNA template. See Scheme 11 for Q and Z designation.

Y = C/T R = A/G
 $\bar{Y} = \frac{m}{C/T}$
 Q = 5'-O-methyl-G
 Z = 2',3'-dideoxy-G



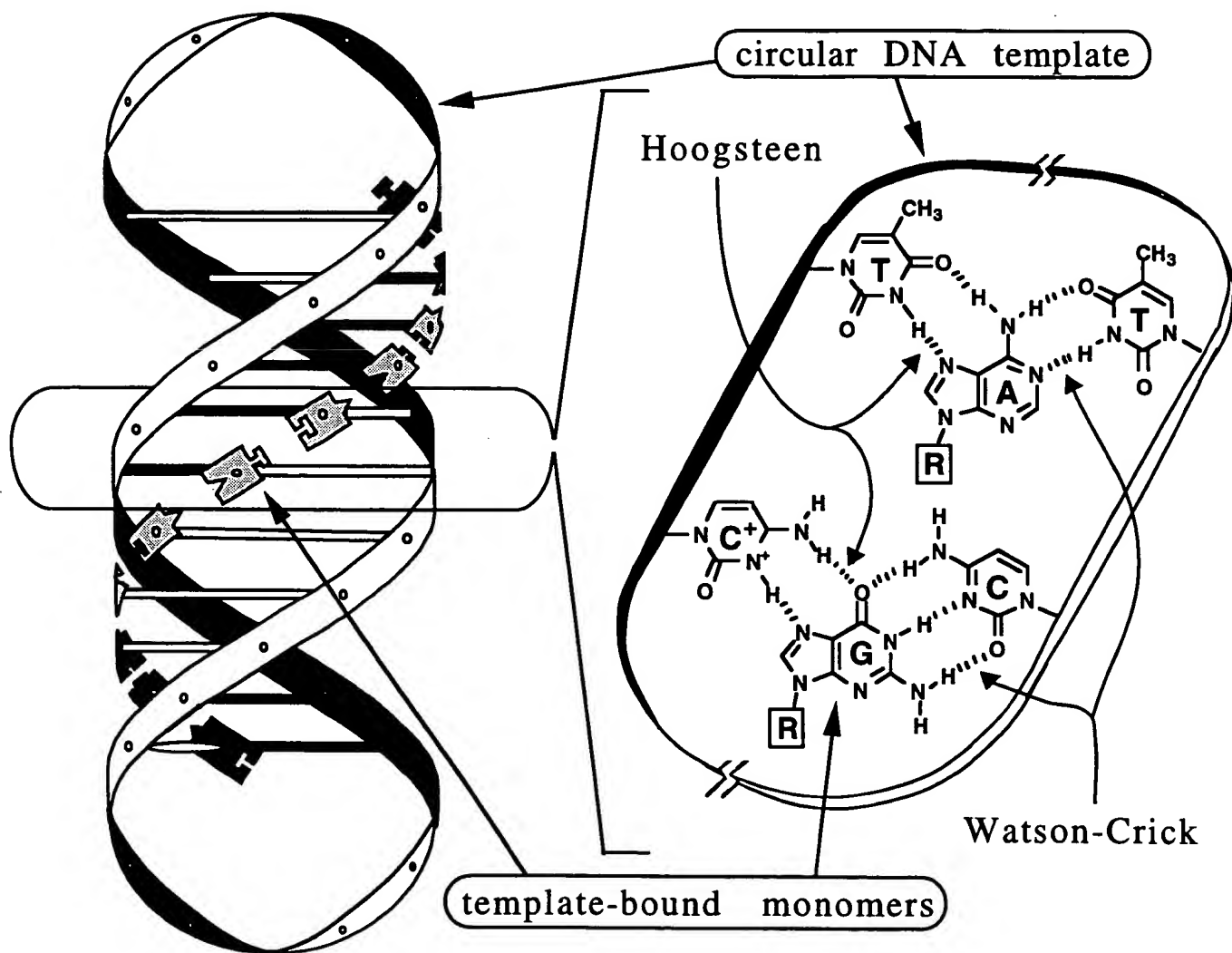


FIG. 7 Ribbon graphic of a circular DNA template with bound monomer nucleobase derivatives and illustration of the template-substrate triplets (**R** represents reacting substrate for oligomerization).

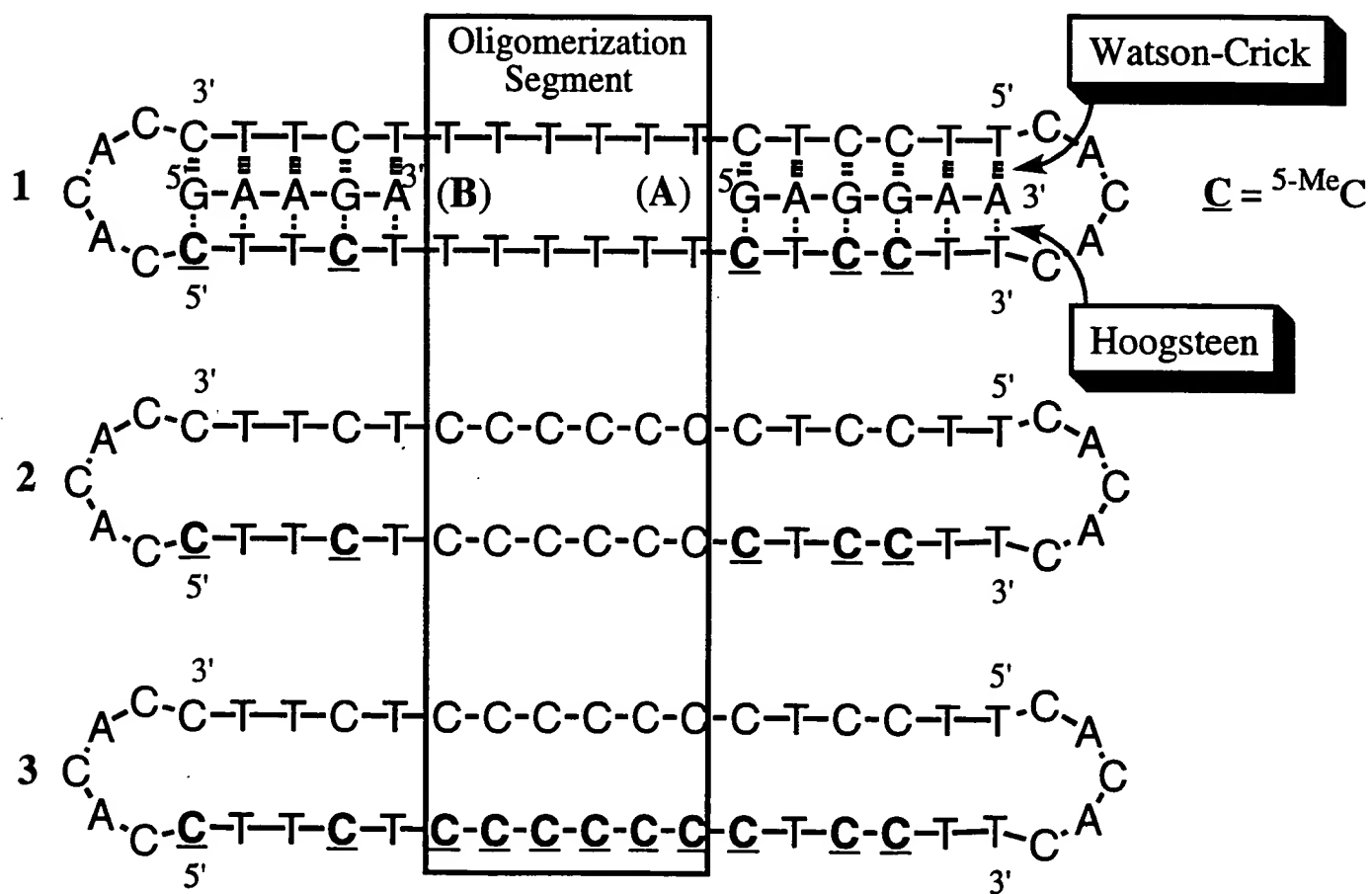
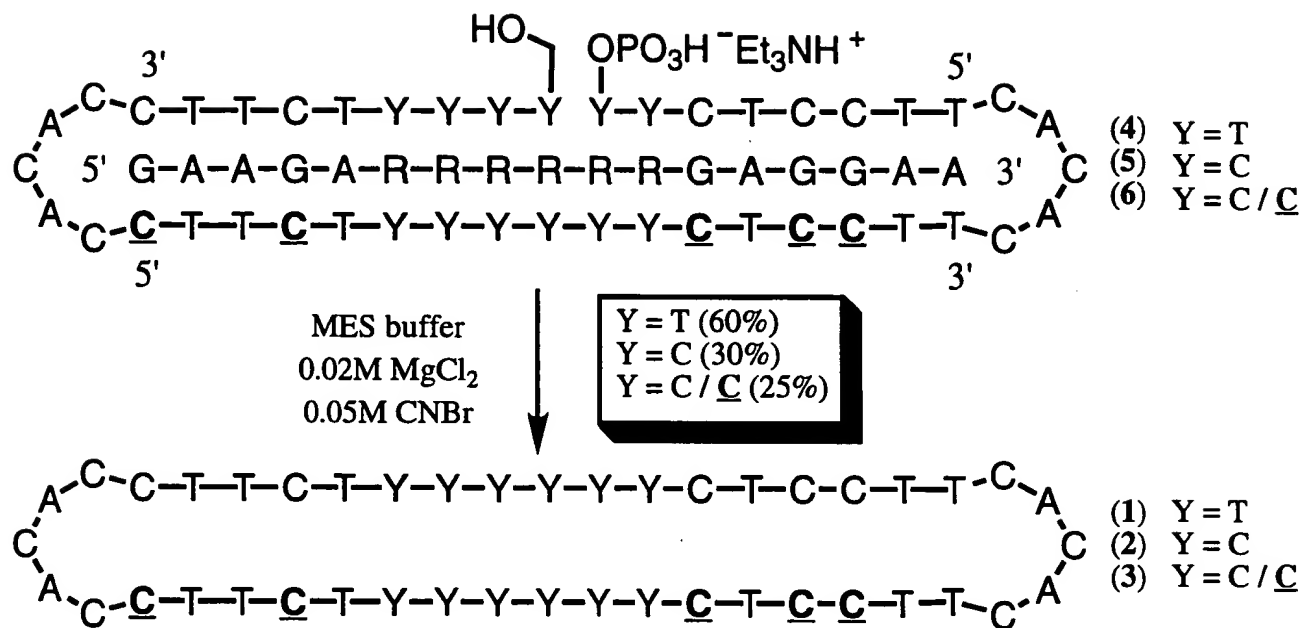


FIG. 8 Three Circular DNA templates (1, 2, and 3) and their respective primers (A and B) for directed ligation and oligomerization experiments. The $\underline{\text{C}}$ designates 5-methyl-cytidine.

Scheme 15



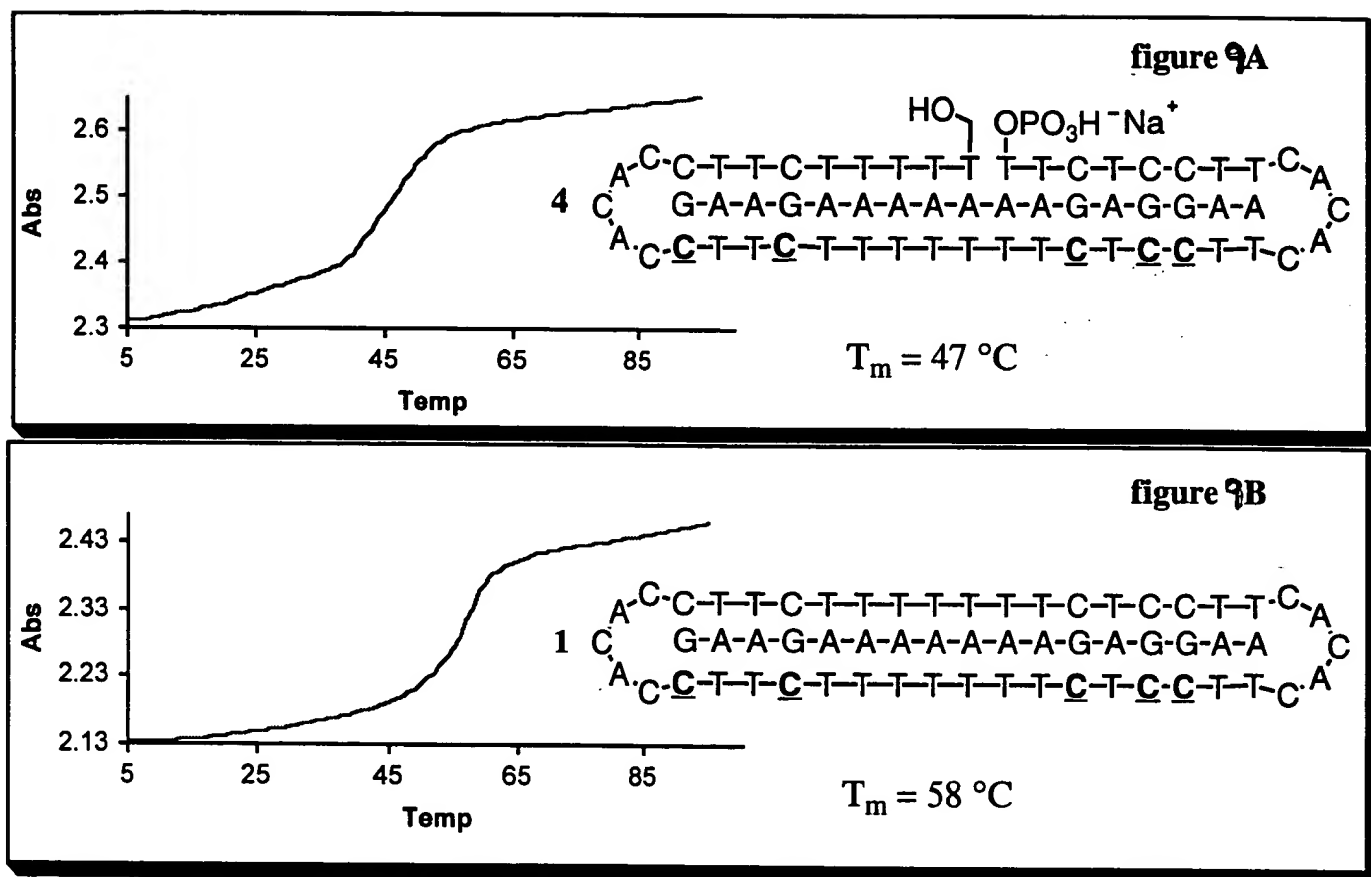


FIG. 9 Melting analysis of 4 with the purine-rich oligonucleotide (3A) and circularized template 1 with the same oligonucleotide (3B).

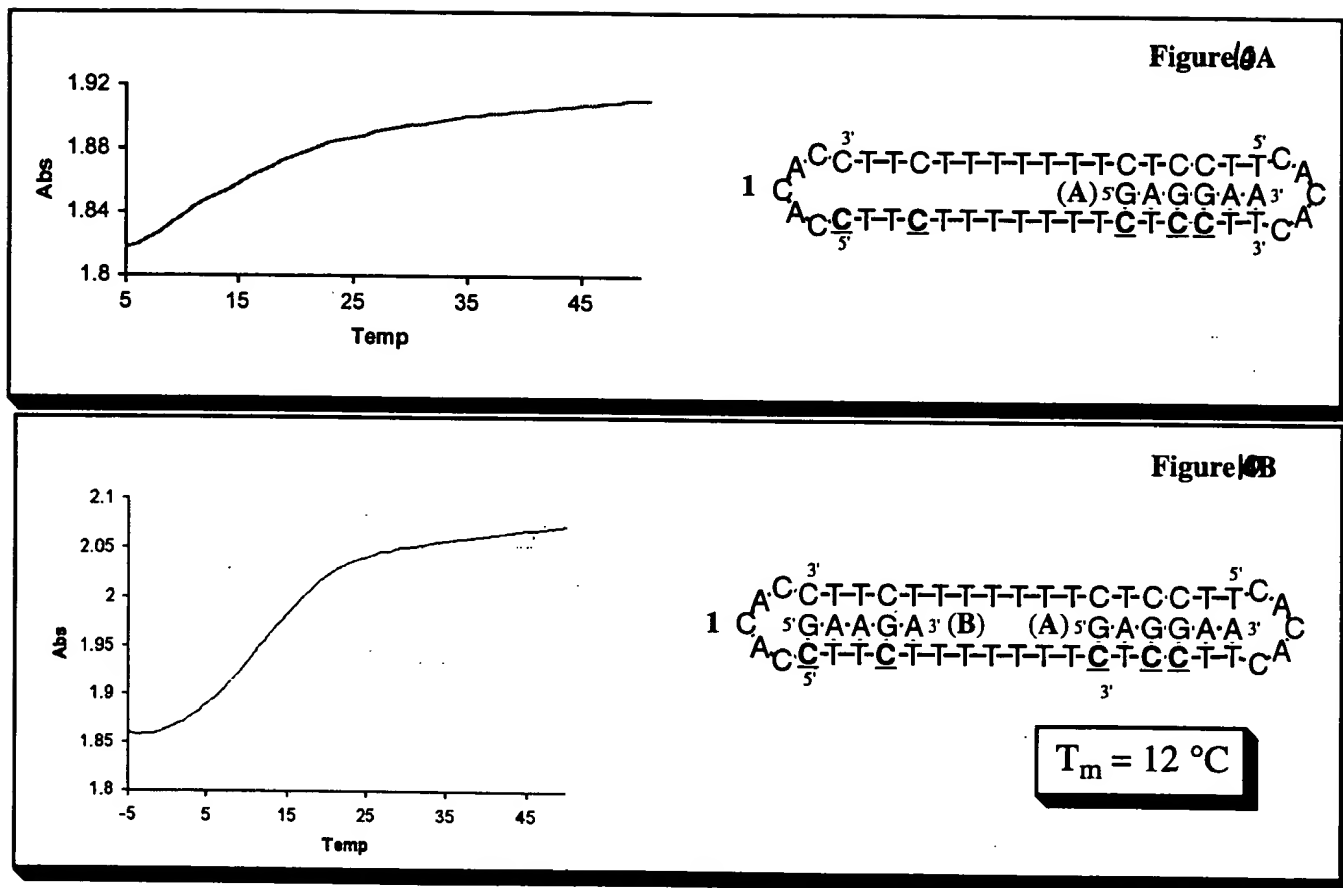


FIG. 10 Melting analysis of circular template 1 with primer A (fig 0A) and primers A and B (fig 0B).

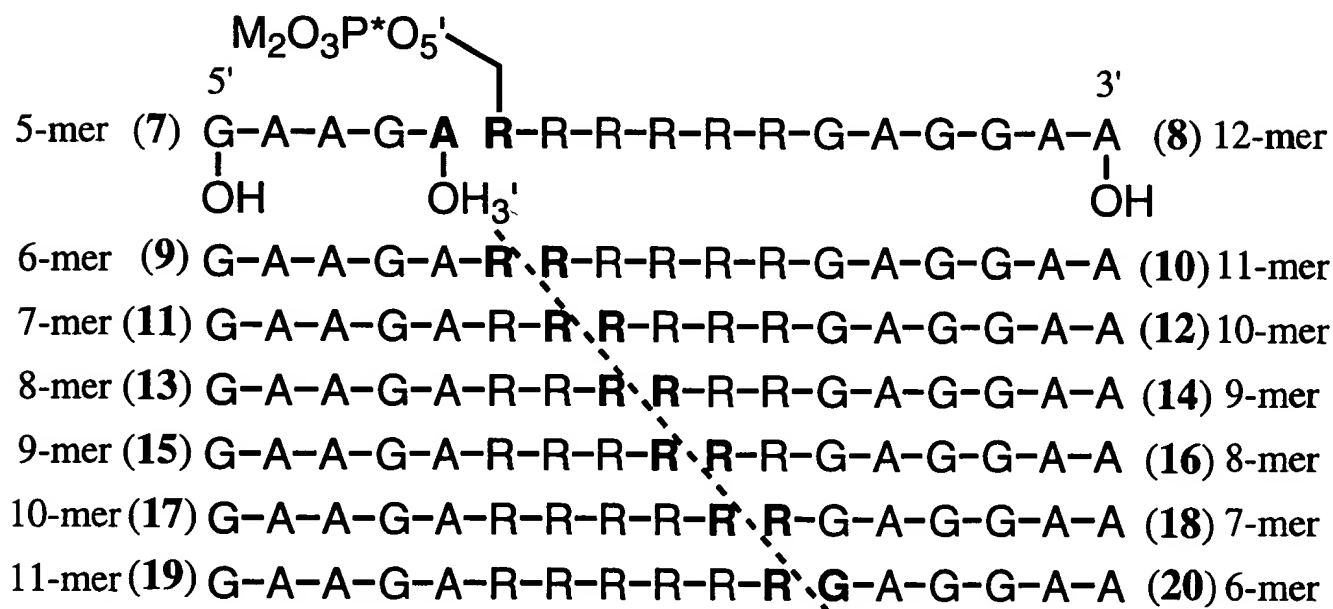
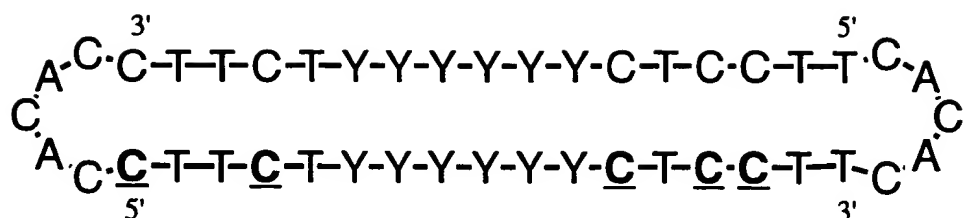


FIG. 11 Ligation analysis investigations in order to probe the effect of Oligonucleotide length and directionality on the ligation reaction. P* represents the P^{32} labelled phosphate which will undergo phosphodiester bond formation.

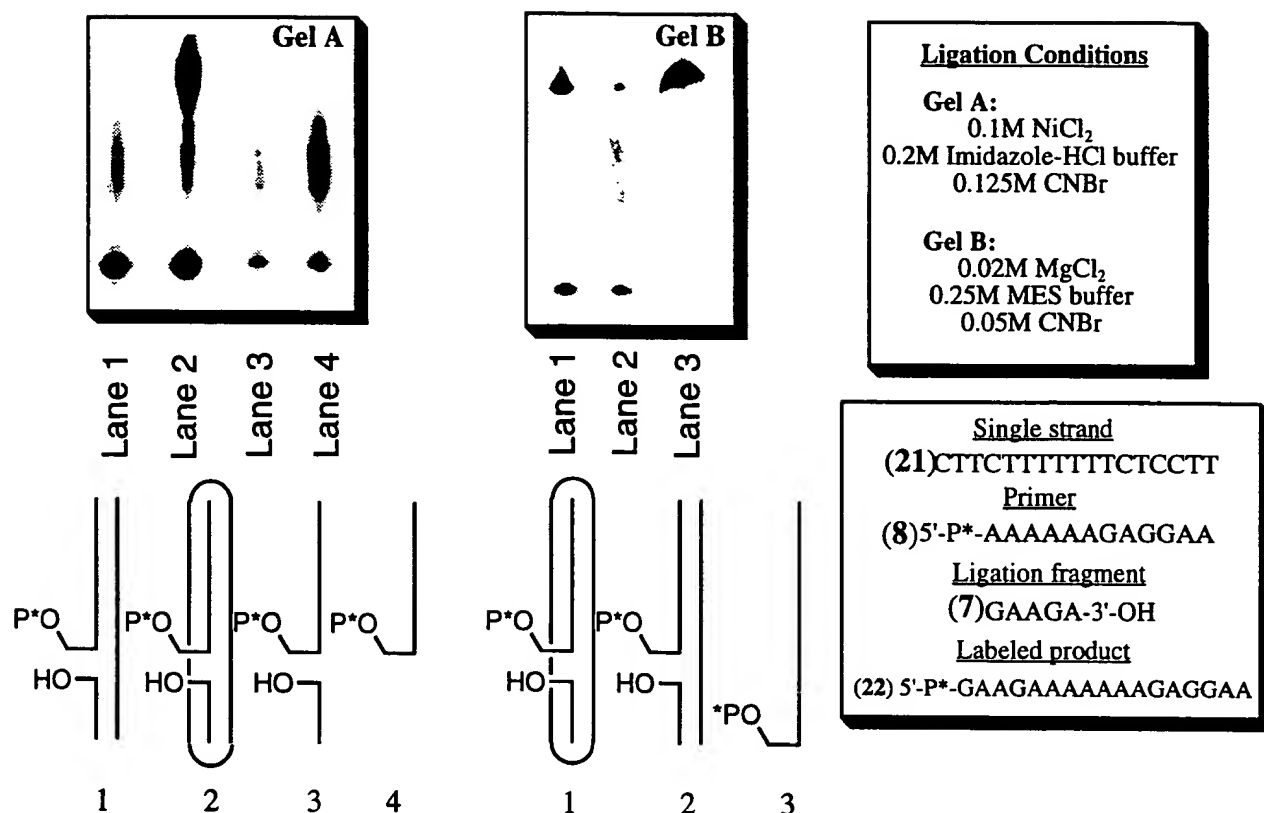
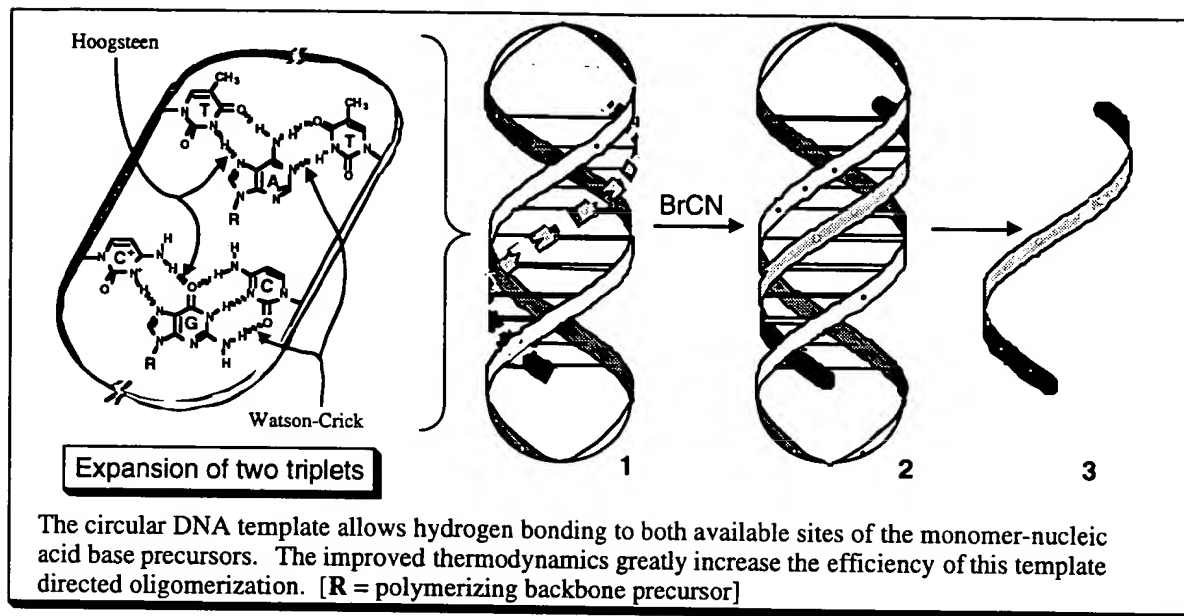
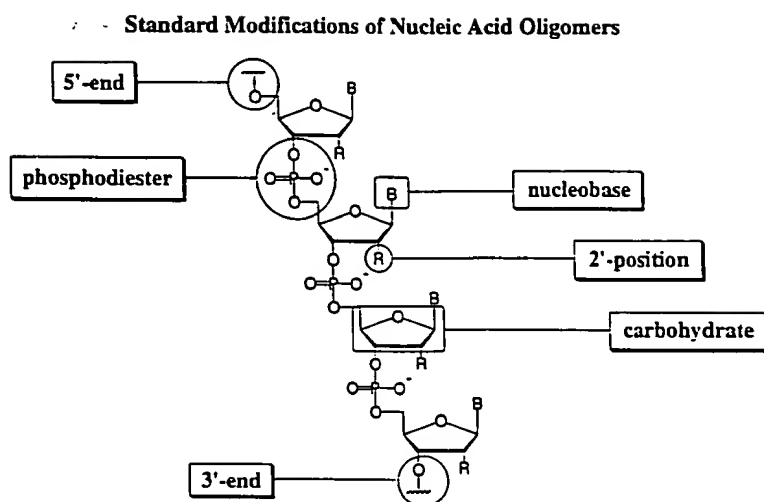


FIG. 12

Autoradiography of PAGE analysis of the ligation of 5-mer 7 with 5'-32 P labeled 12-mer 8. Gel A shows the results of the reaction run with NiCl₂ in imidazole•HCl buffer, while Gel B shows the result of the reaction run with MgCl₂ in MES buffer. Lanes 2 (Gel A) and 1 (Gel B) show the migration of the reaction mixture from ligation on circular DNA template 1. Lanes 1 (Gel A) and 2 (Gel B) compare the results of the ligation reaction on the corresponding single-strand DNA template X. Lane 3 (Gel A) shows the results of the ligation reaction with no template present. Lane 3 (Gel B) shows the migration of the expected full length ligated product which was independently synthesized and 5'-32 P labeled.



Scheme 16



- (10) Beaucage, S.L.; Iyer, R.P. *Tetrahedron* 1993, 49, 1925-63.
 (11) Uhlmann, E.; Peyman, A. *Chem Rev.* 1990, 90, 543-84.

Figure 13. Components of nucleic acid polymers that are commonly modified to induce selective properties or functionality to an oligomer.

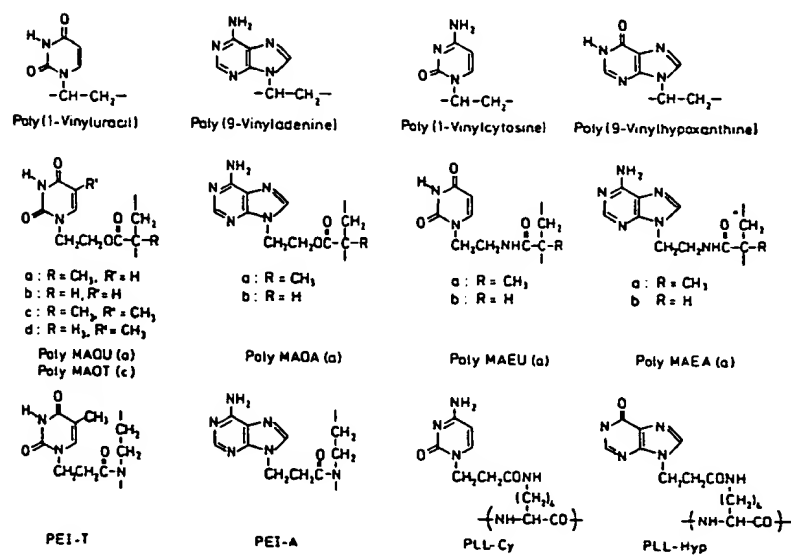


Figure 14. Synthetic Nucleic Acid Analogs.